

# Complete genome sequencing of embryos

**Santiago Munné**



# Affiliations and Disclosures

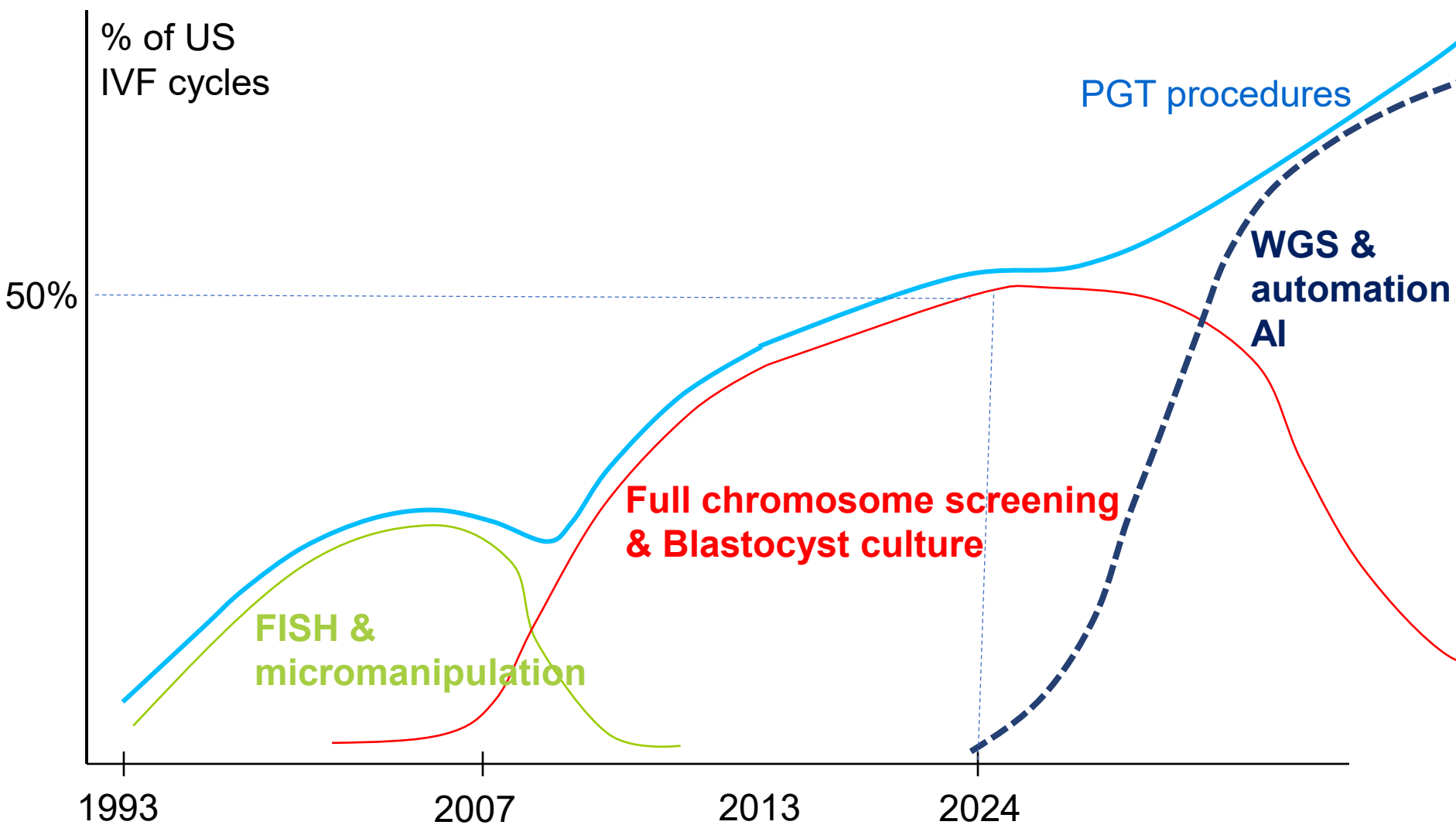
Scientific Director @ **Progenesis**

President, founder @ **HoMu Health Ventures**

Chief Innovation Officer, founder @ **Overture Life**

BOD @ **GenEmbryomics, Butterfly Bio, Sama, Ovum Health**

# Waves of technology



Morphology

FISH

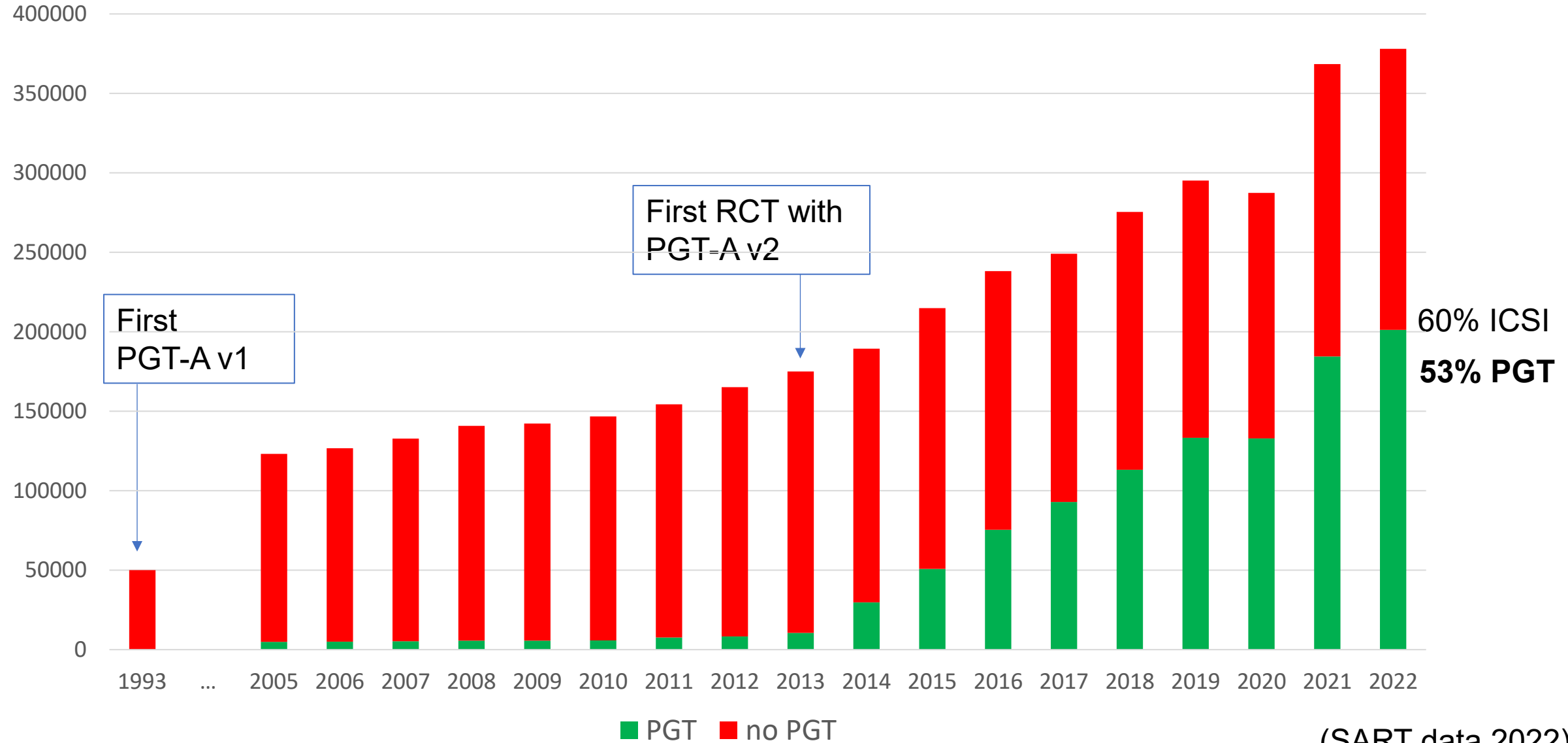
CGH

NGS

WGS

# PGT usage: in the US is the standard of care

US cycles



(SART data 2022)

# My journey through PGT

**Genetics  
before PGT:**  
Not actionable

1993 - 2012

**PGT-A v1:**  
Actionable  
Needed skills  
More depth

2013- 2020

**PGT-A v2:**  
Genetics automated  
IVF and Biopsy was not  
PGT-A is boring



2020-2025  
**PGT-WGS, PGT-P**  
Fun again  
Not enough eggs

TBC

2025

**Lab automation  
/ non invasives:**  
IVF lab automated  
Non-invasives still need work

# Comprehensive PGT methods

## SNP ARRAYS GENOTYPING

- Karyomapping (1)
- Haplarithmisis (2)
- High throughput SNP array (13)
- HaploSeq
- APCAD (4)
- Genome prediction PGT-PS (3)
- Haplotype-Aware (5)
- Whole genome prediction (6)

## GENOTYPE BY SEQUENCING

- PGT Complete (12)
- Targeted NGS
- One PGT (7)
- Genotyping by sequencing (8)
- Chen et al. (9)
- HaploPGT (10)
- S-HaploSeek (11)

## SEQUENCING + ADDING SNPs

- PGT-Seq
- OneGene PGT (14)

(1) Handyside et al. *J. Med. Genet.* 47, 651–658 (2010); (2) Zamani Esteki, et al. , *Am. J. Hum. Genet.* 96, 894–912 (2015). (3): Treff et al. (2019) *Frontiers Endocrinol* (4) Verdyck et al. 2022, (5) Ariad et al. (2021) . (6) Kumar (2022) *Nature Medicine* 28:513–516, (7) Masset et al. *Hum. Reprod.* 34, 1608–1619 (2019); (8) Masset et al. (2022) *Nucleic Acid Res*, (9) Chen et al. (2020) *Human Reprod*; (10) Xie et al. (2022) *Human Reprod* 37:2546–59.(11) Backenroth et al. (2023) *Nature Scientific Reprots* 13:18036 (12) Buldo-Licciardi et al. 2020. *ASRM*; (13) Treff et al. *Eur J Med Genet.* 2019;62: 103647. (14) Hornak et al(2024)

# Genotyping: High throughput SNP arrays

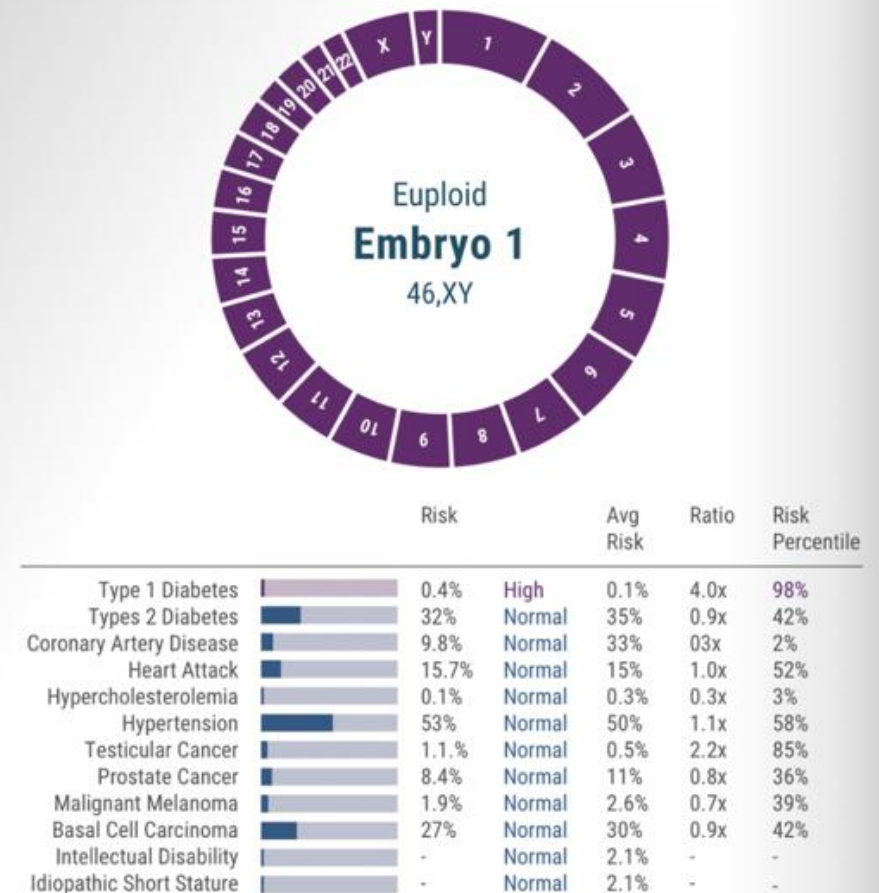
High throughput SNP microarray (800K SNPs)

**PGT-A: quantitative** copy number analysis and **qualitative** by genotyping: with **99.3% accuracy** <sup>(1)</sup> and detects **all polyploidies** and UPD <sup>(2)</sup>

**PGT-M:** using **parental DNA** it does **linkage analysis** and detects 20-500 SNPs within 2M window of the gene defect

**PGT-SR:** differentiates **Normal from Balanced**, with a 10Mb resolution <sup>(1)</sup>

**PGT-P:** for several diseases, with AUC of 0.65-0.75



(1) Treff et al. (2019) Frontiers Endocrinol.; Leahy et al.. JARG 41: 121-126; (2): (2) Kratka et al. (2023) F&S science, **Diego Marin in da room**

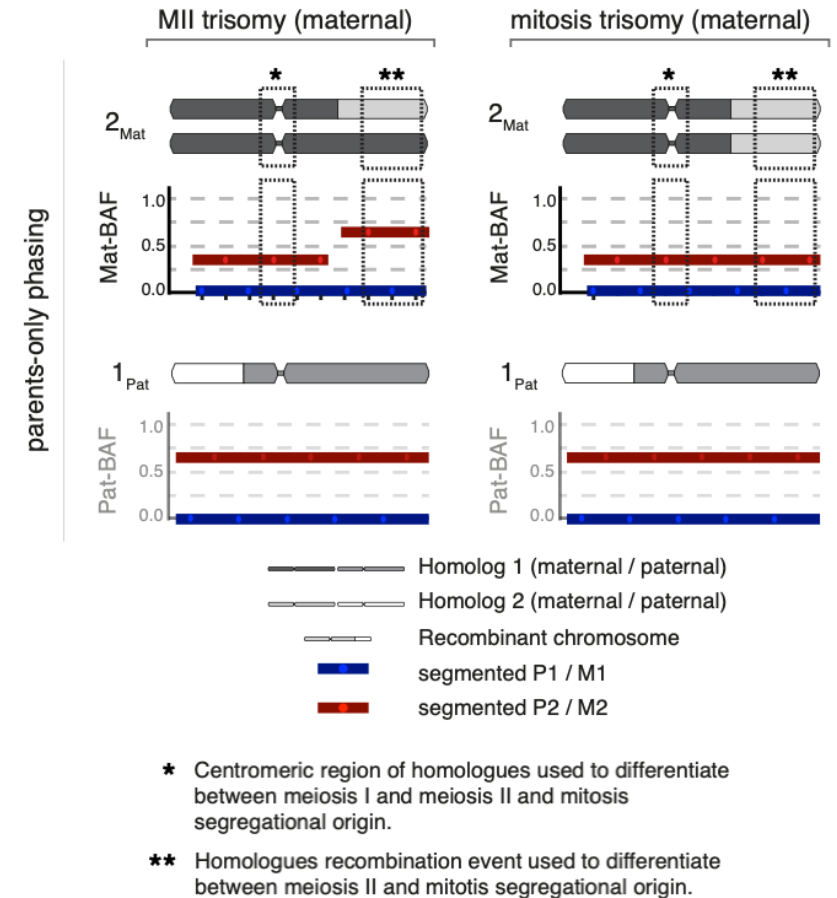
# Genotyping by sequencing: Haplorithmisis

## METHOD:

- MDA amplification
- Parental and embryo DNA is **sequenced at x10 depth**
- haplotyping analysis using **HAPLORITHMISIS**

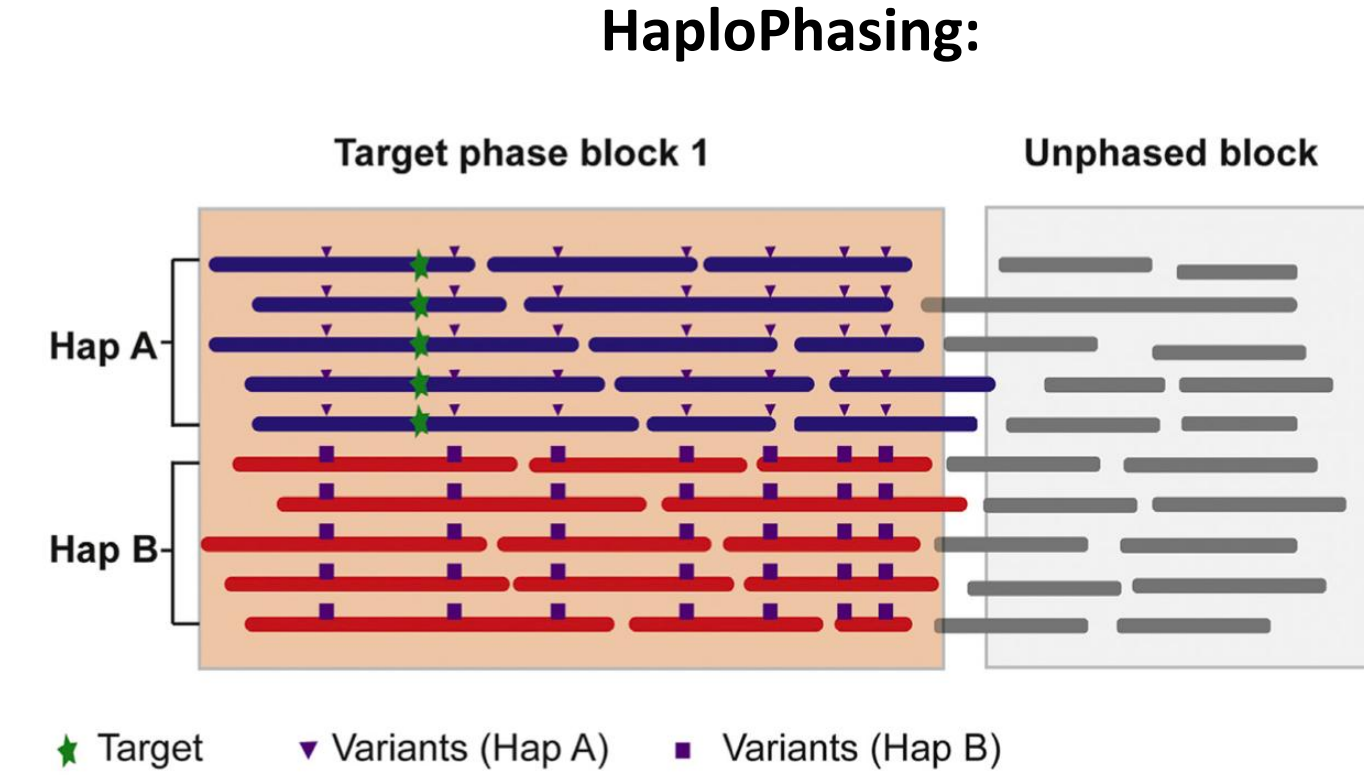
## RESULTS:

- 80% genome coverage
- **PGT-M: direct mutation for 82%** embryos and rest by haplotyping
- **PGT-A: Detects mitotic origin** of mosaicism, all **triploids**
- **PGT-SR: differentiates balanced from normal**



# HaploPhasing by Long Read sequencing

- **Long read sequencing** of the **parents** provides parental variant maps
- **Tiling of variants** allows SNP phasing around the target sequence.
- **PGT-A**: standard method
- **PGT-M and PGT-SR**: through SNP PCR for mutation or or breakpoint. No proband needed



NextSeq, 8M reads.

Cheng et al. (2021) *Fertil. Steril.* 116:774-783

# Why do WGS?

***De novo* mutations** are not detectable in the parents.

# Rare Diseases and de novo mutations

- There are **20,000 genes** coding for proteins
- **10,000 genetic diseases** have been identified, of which **>7000 are rare**<sup>1</sup>
- The basis for **these rare diseases** can be identified by **whole exome sequencing**<sup>2</sup>:
  - 32%** in non-consanguineous patients
  - 50%** in consanguineous patients
- **300 million people** worldwide have a **rare disease**<sup>3,4</sup>
- Over **80% of severe developmental disorders** are caused by ***de novo* mutations**

# 40% more congenital abnormalities in ART

IVF babies have **40% more congenital abnormalities** than naturally conceived babies.

Reasons:

- **higher parental age** of IVF patients
- **underlying mutations** causing infertility.

ginal Sound for Musicians: Off ● Recording



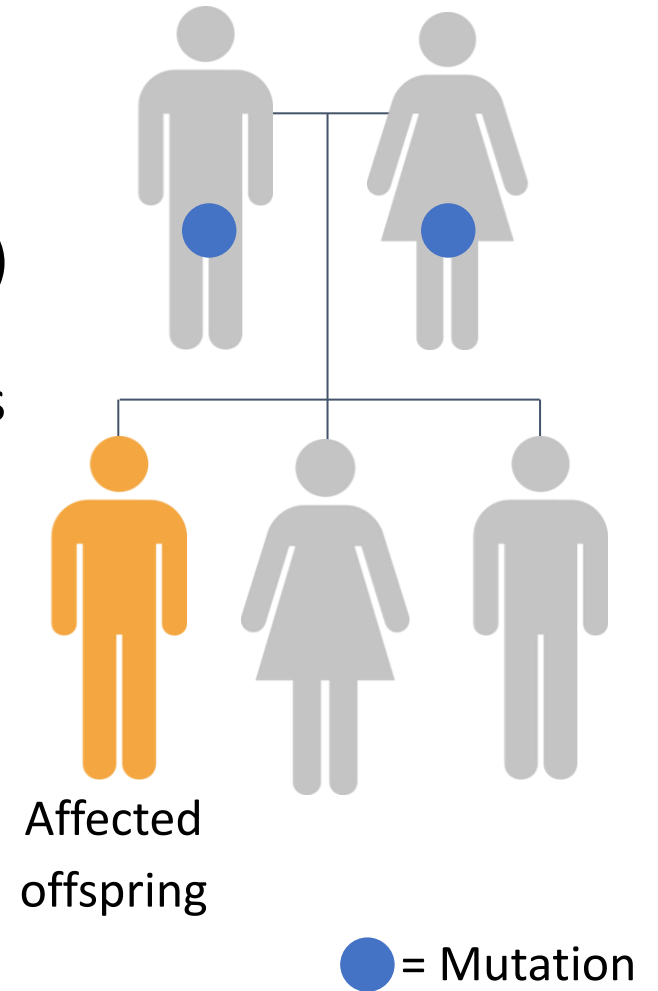
## Congenital anomalies ART vs natural

Meta-analyses	Articles	ART infants	Pooled estimate (95% CI)
Wen <i>et al</i> (2012)	46	124,468	1.37 (1.26, 1.48)
Hansen <i>et al</i> (2013)	45	92,671	1.32 (1.24, 1.42)
Zhao <i>et al</i> (2018)	46		1.40 (1.31, 1.49)
Hoosnan <i>et al</i> (2017)	30		1.99 (1.87, 2.11)
- Cardiac defects			1.43 (1.27, 1.62)
- CNS defects			1.36 (1.10, 1.70)
- Urogenital defects			1.58 (1.28, 1.94)
- Musculoskeletal			1.35 (1.12, 1.64)
- Chromosomal			1.14 (0.90, 1.44)
Giorgione <i>et al</i> (2018)	8	25,856 CHD	1.45 (1.20, 1.76)
Zhao <i>et al</i> (2019)	46	112,913	1.43 (1.31, 3.52)

**ARTs increase risk of birth defects by about 40%**

# De novo mutations (DNV)

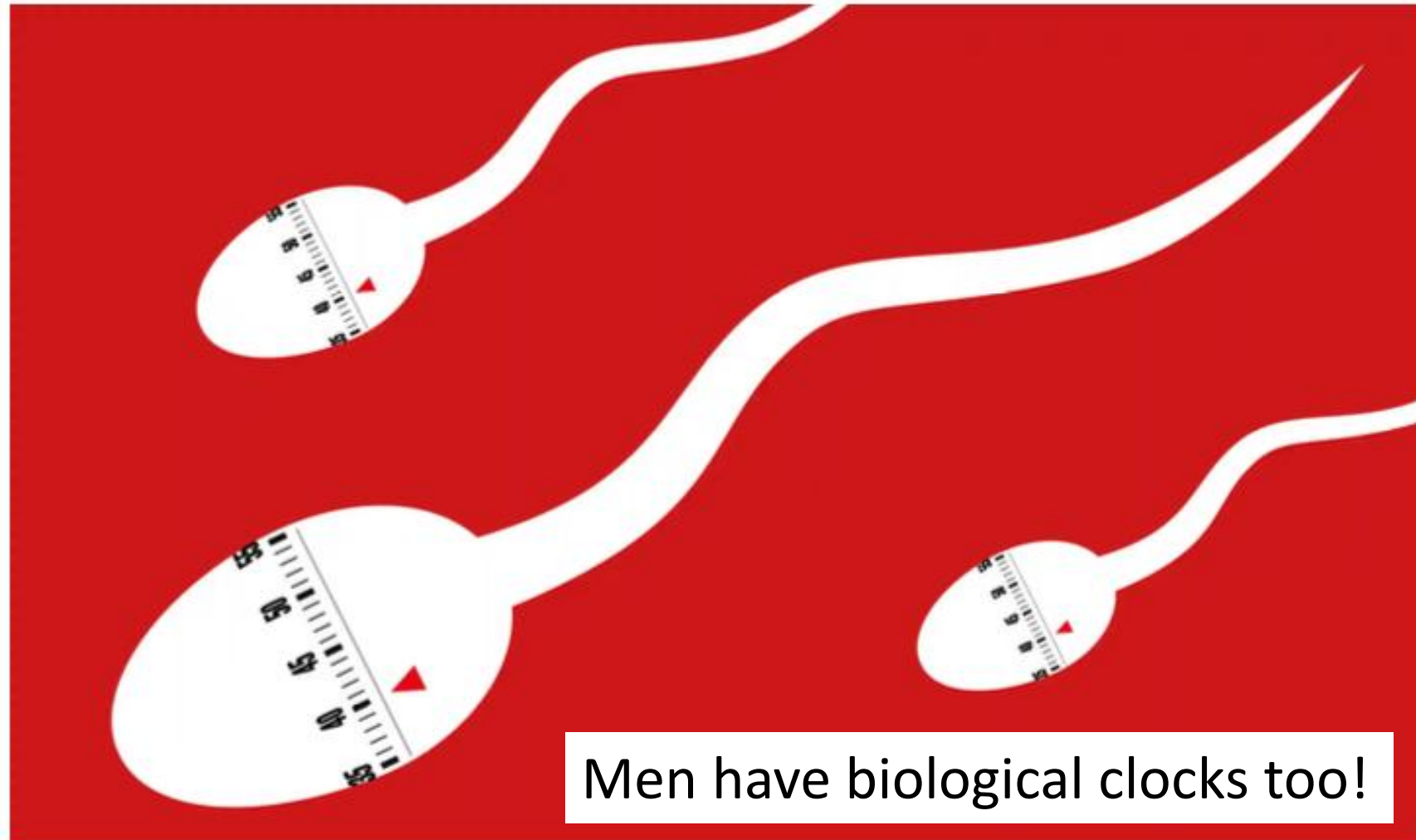
- DNV mutations accumulate in the parents' gametes and are **not detectable by carrier screening**.
- There are **74 DNV mutations per embryo: 1-2 pathogenic** (1-3)
- DNV mutations are **more detrimental** than inherited mutations not exposed to natural selection (11)
- DNV mutations **increase with paternal age** (4).  
Fathers 45 years old have x3.5 risk of autistic (5,6) and x27 risk of bipolar offspring than 25 years old (7).
- **1/488 children are born with a DNV mutation causing developmental malformation** (8,9), Autism (10), ....



(1) Acuna-Hidalgo et al. (2015) *Am J Human Genet* 97, 67–74, (2) Kondrashov (2003) *Human Mutation* 21, 12–27, (3) Acuna-Hidalgo et al. (2016) *Genome Biology* 17, 241. (4) Kong et al. 2012, *Nature*, (5): D’Onofrio et al. 2014, *JAMA* (4) Sanders *Nature* 2012;485:237–41.(7) Sandin S et al. (2016). *Mol Psychiatry*. 21:693–700 (8) *Lord Lancet* 2019;393:747–57, (9) de Ligt *N Engl J Med* 2012;367:1921–9. (10) Kong et al. 2012, *Nature*; (11): Veltman *Nat Rev Genet* 2012;13:565–75.

# The perils of putting off fatherhood: why it poses risks to children's physical and mental health

The  
Guardian

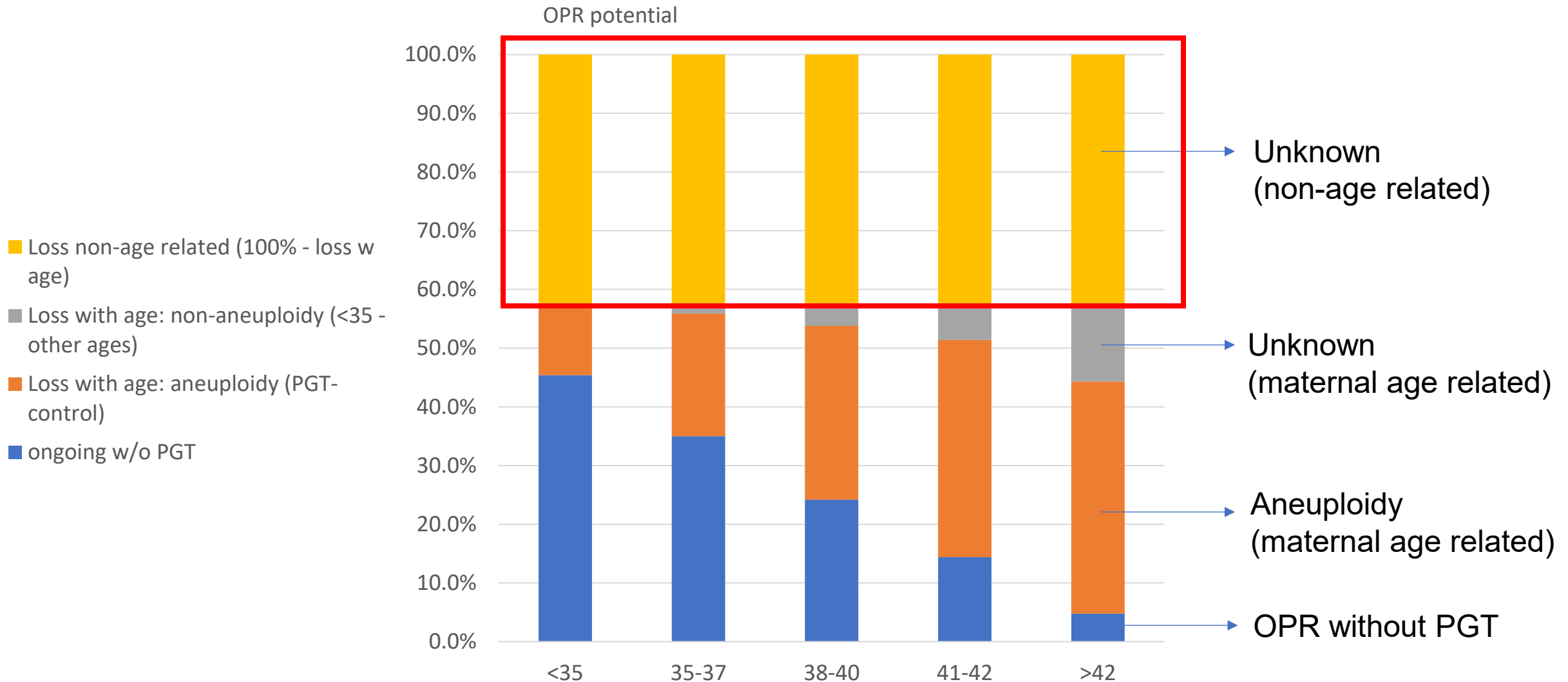


Men have biological clocks too!

Older men are not generally discouraged from using fertility services, unlike women. Illustration: Philip Lay/Observer Design

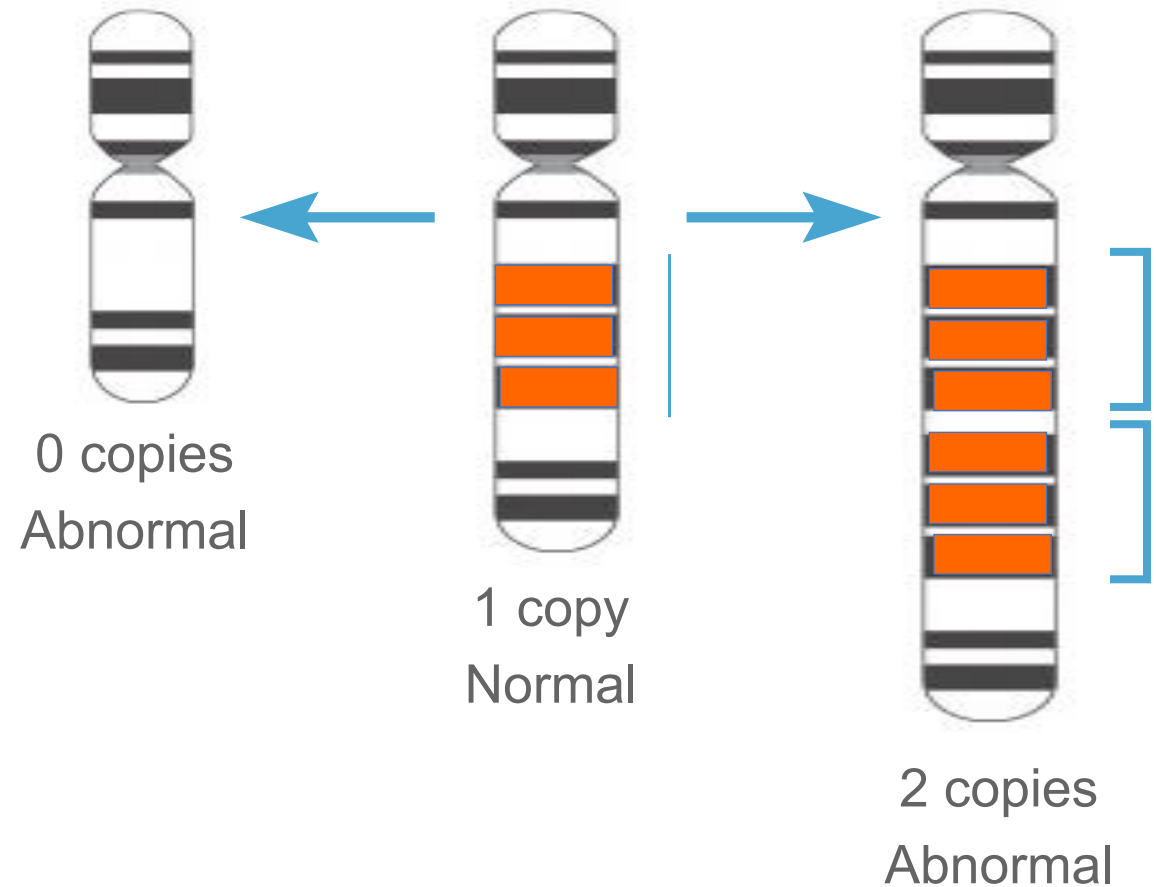
Men have biological clocks too. Fertility drops with age, and the likelihood of offspring having conditions such as autism, schizophrenia and leukaemia rises

# De novo mutations could produce embryo lethality

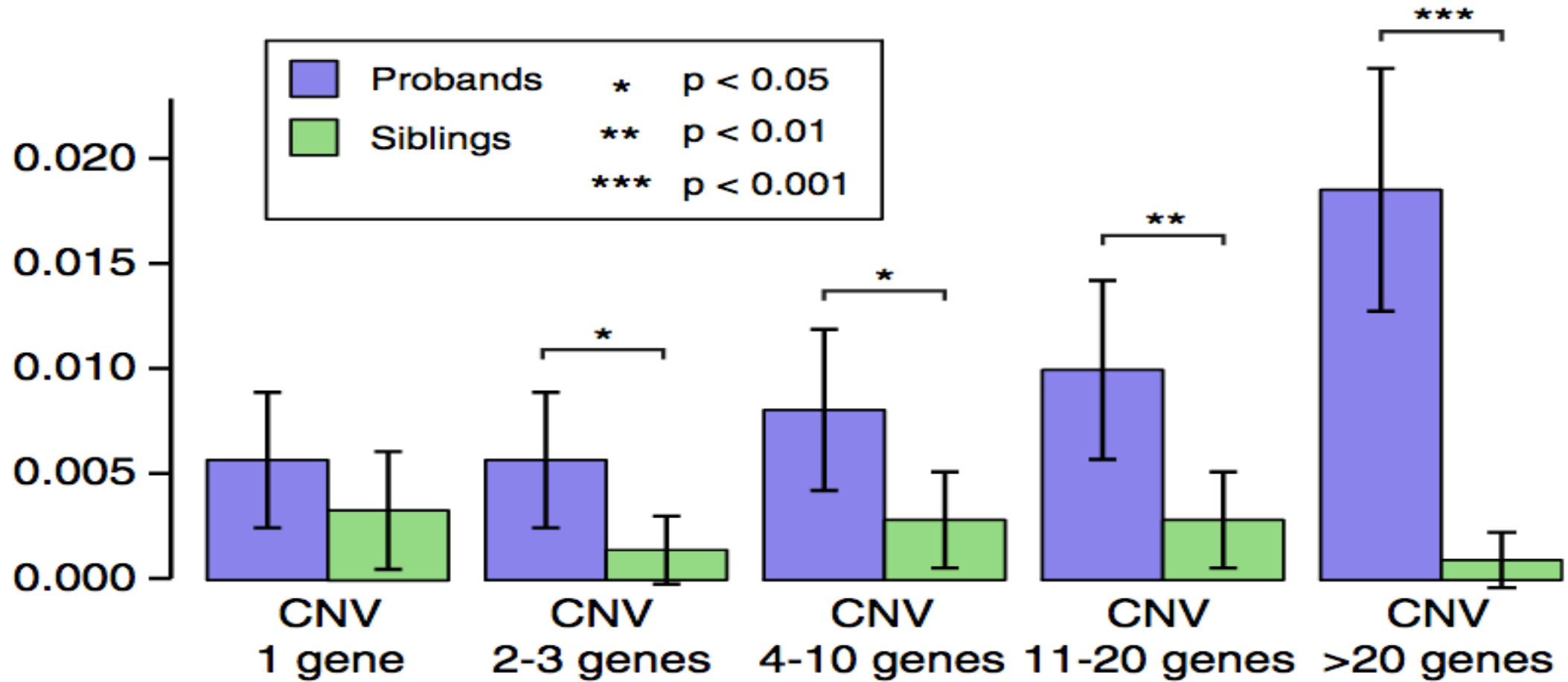


# De novo copy number variants (CNV)

- **Copy number variants (CNV)** are the result of segmental duplications and /or deletions >500bp
- Unlike point mutations, **CNVs are extremely unlikely to be amplification artifacts**
- **1-2% of conceptions** carry CNVs that are 100kb to 10Mb, not detectable by low-coverage PGT-A



# High number of CNVs is very predictive of autism



# PGT methods for Whole Genome sequencing

1. **Pieters, Munné et al (2015):** Individually sequence 384 DNA fragments to allow haplotyping to detect amplification artifacts.



2. **Xia et al. (2021):** Uses PTA to amplify the DNA biopsy



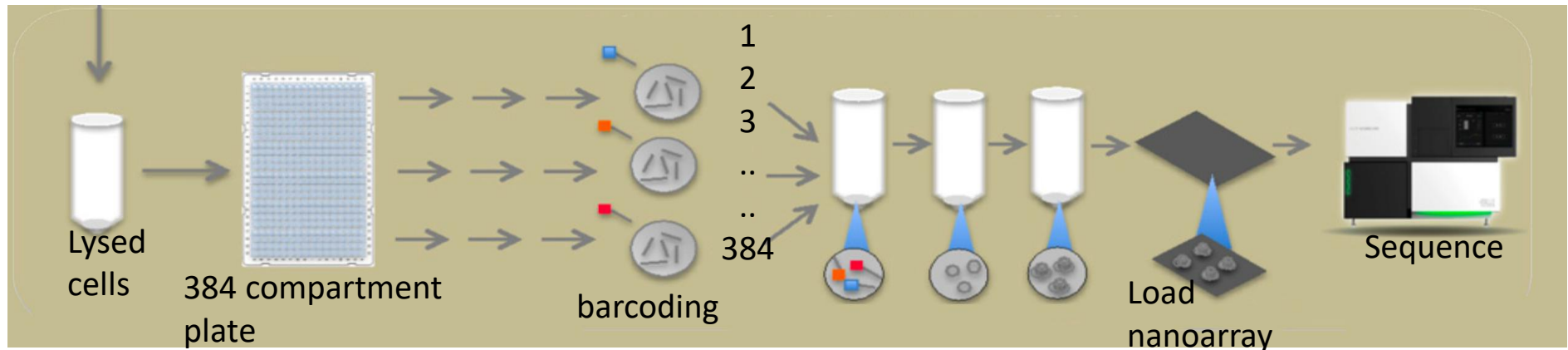
3. **Murphy et al. (2020):** standard amplification with bioinformatics filtering of false positives



# 1. WGS using Large Fragment Reads (LFRs) (Pieters et. al 2015)

## METHOD:

- DNA was dispersed across **384 compartments**, each with 5% haploid genome
- Amplification, fragmentation, barcoding, and sequencing at x23 depth, **were performed individually for every compartment**



## RESULTS:

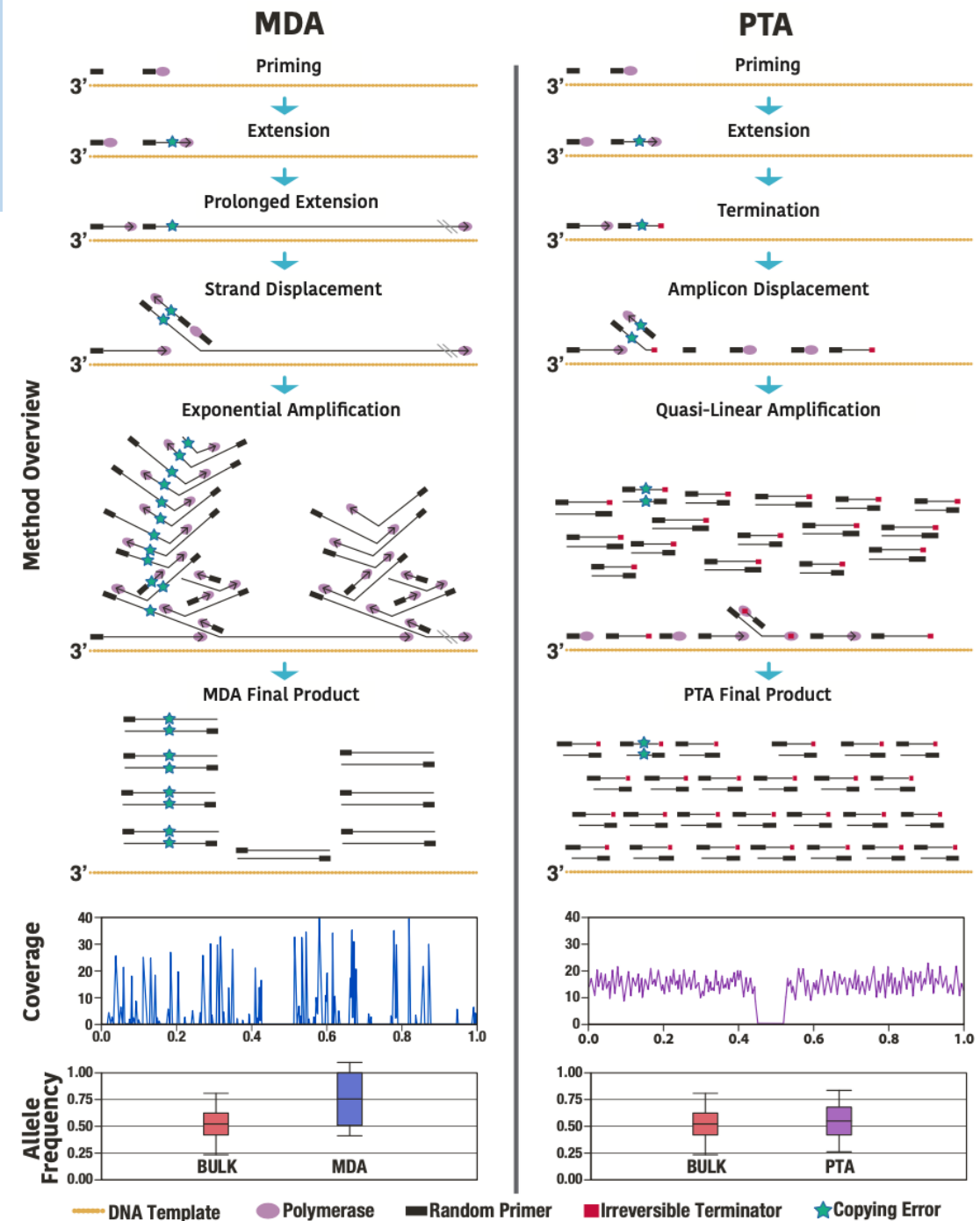
- **97%** of the biopsy genome was sequenced
- **3000 de novo variants** per embryo, mostly **VUS**, but **30 pathogenic**

## 2. Primary template-directed amplification (PTA)

### PTA method:

- Uses MDA polymerase but **exonuclease-resistant terminators** that create smaller amplicons.
- The amplicons undergo **limited subsequent amplification** with more of the amplification occurring from the primary template
- **Any errors** in daughter amplicons have **limited propagation** during the following amplification

*Gonzalez et al. (2020), Xia et al. (2021)*



## 2. PTA: Results

# ORCHID

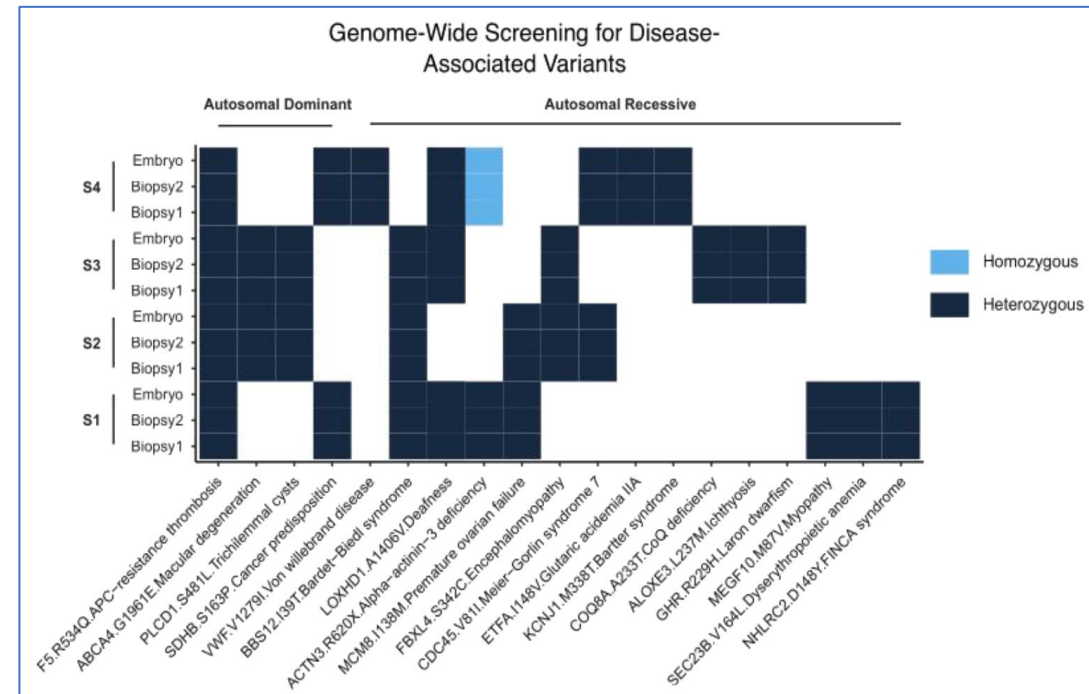
**PGT-A: 99% concordance** for chromosome abnormalities in 110 reanalyzed embryos<sup>(2)</sup>.

**PGT-SR: CNVs greater than 400Kb** were detectable.

**PGT-M: no need of parental DNA at x30 depth**

**7-9 pathogenic variants per embryo<sup>(1)</sup>:**

- 11 inherited (appearing in >1 embryo).
- 7 De novo (appearing only in one embryo)



**Detected 3.27M SNVs, 96% concordant with the whole embryo (70,000 SNVs different)<sup>(1)</sup>**

- Uses **PTA** amplification which provides **better coverage** of the genome
- Reporting on **1200 genes**
- **Offered clinically** with over 250 performed procedures (\$2500 per embryo)
- They **do not test the parents**:
  - Cannot **confirm the mutation** using haplotype phasing from parental DNA
  - Cannot **differentiate de novo mutations from artifacts**

### 3. GenEmbryomics: method



#### Method:

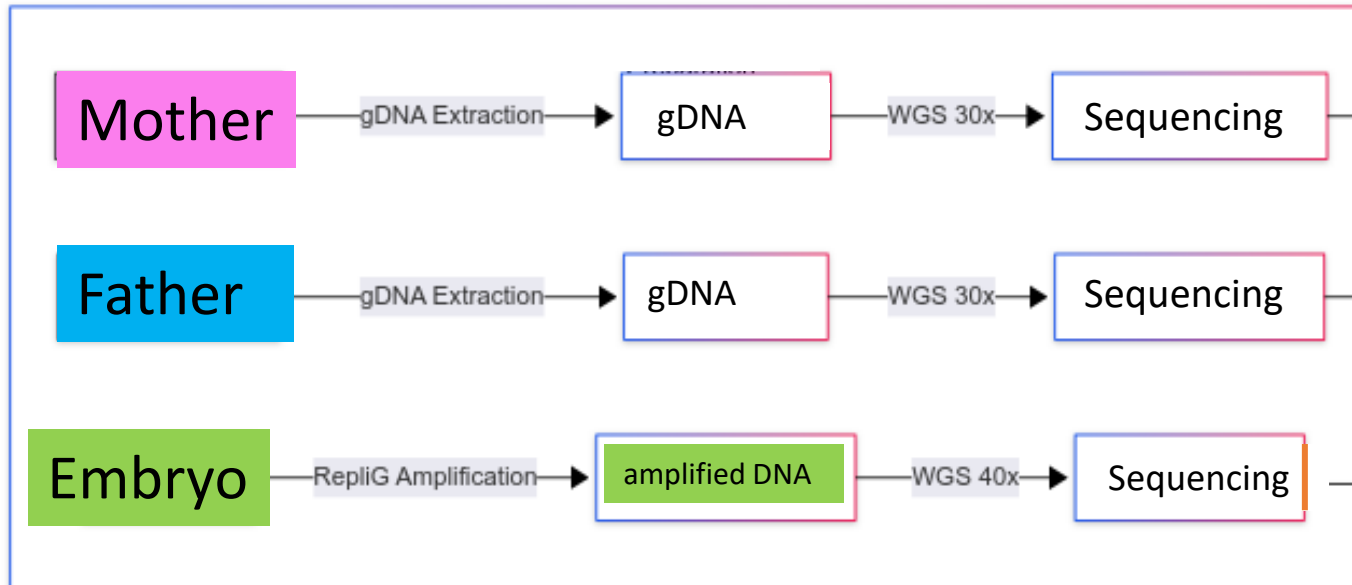
- **WGA** by MDA
- **Whole genome sequence** of parents at **x30** and embryos at **x40 depth**
- **Extensive Variant filtration** to eliminate false-positives
- **Variants annotation** is performed from >50 annotation sources, pathogenicity prediction algorithms, and ACMG guidelines.

#### Outcomes:

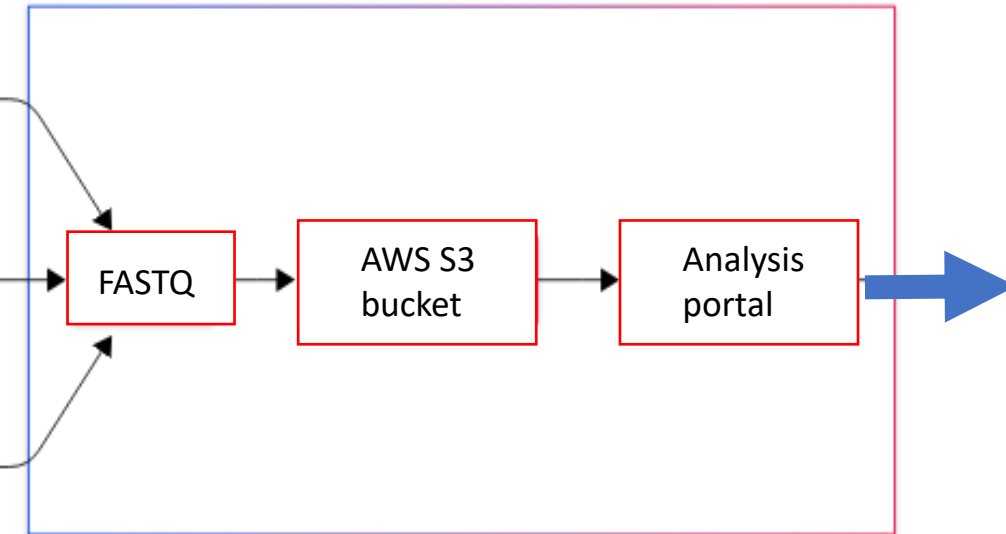
- **Inherited** mutations
- ***de novo*** mutations
- Aneuploidies, translocations, triploidy, mosaicism, Copy number variants >10Kb
- Carrier **trinucleotide repeats**
- **Polygenic** gene disorders and traits

# Pipeline Part 1: Sample Collection & Data Transfer

## SAMPLE COLLECTION AND PREPARATION

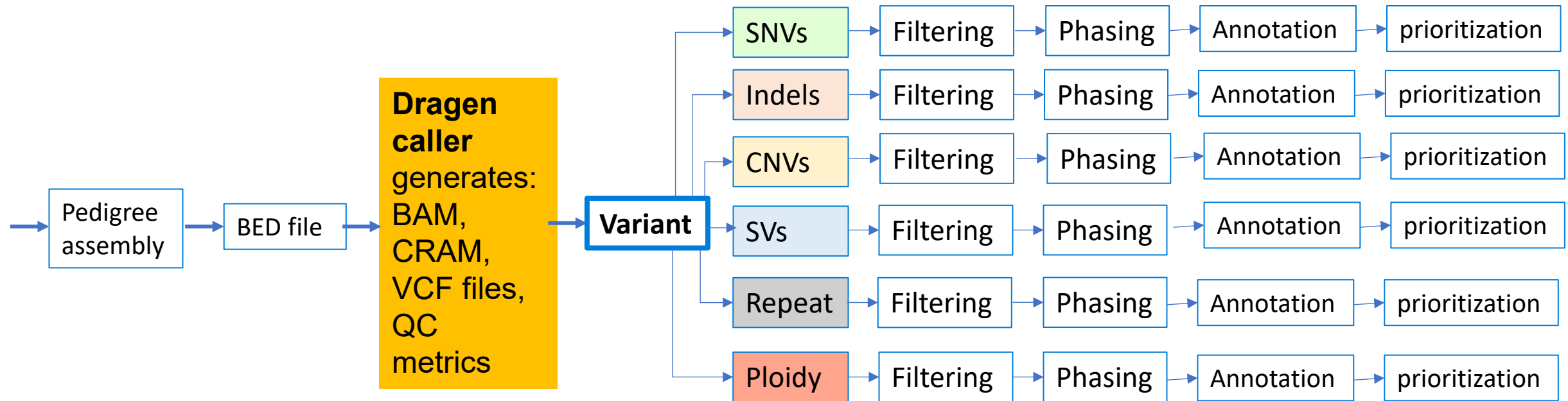


## DATA TRANSFER AND STORAGE



- **Mother and father** samples undergo gDNA extraction followed by WGS sequencing at x30
- The **embryo biopsy** is **amplified** by RepliG followed by WGS sequencing at x40
- Sequencing data is **converted to FASTQ format** for analysis

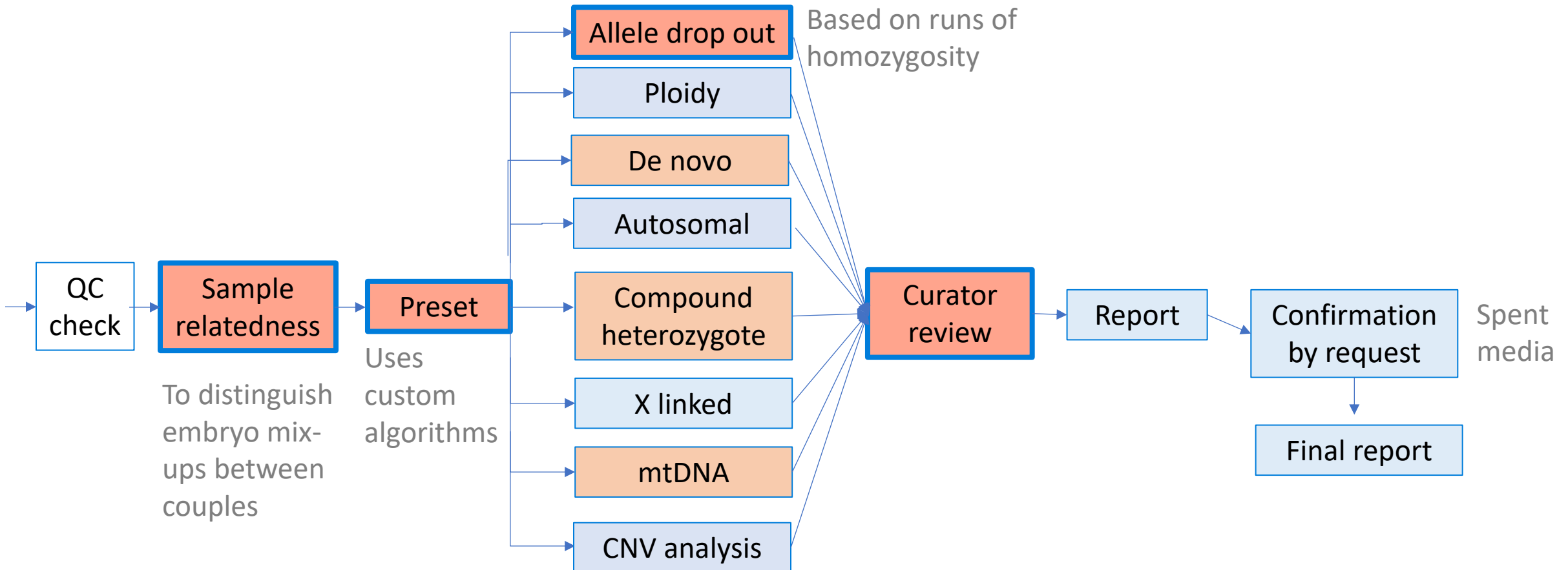
# Pipeline Part 2: Analysis Pipeline



- **Pedigree** is assembled and converted to a **BED file**.
- The **Dragen** calling suite **generates** VCF, CRAM, BAM files and QC metrics
- **Variant calling branches into** SNVs, Indels, CNVs, SVs, Repeat analysis, and Ploidy calling.
- All variants are then **filtered, phased, annotated, and prioritized**

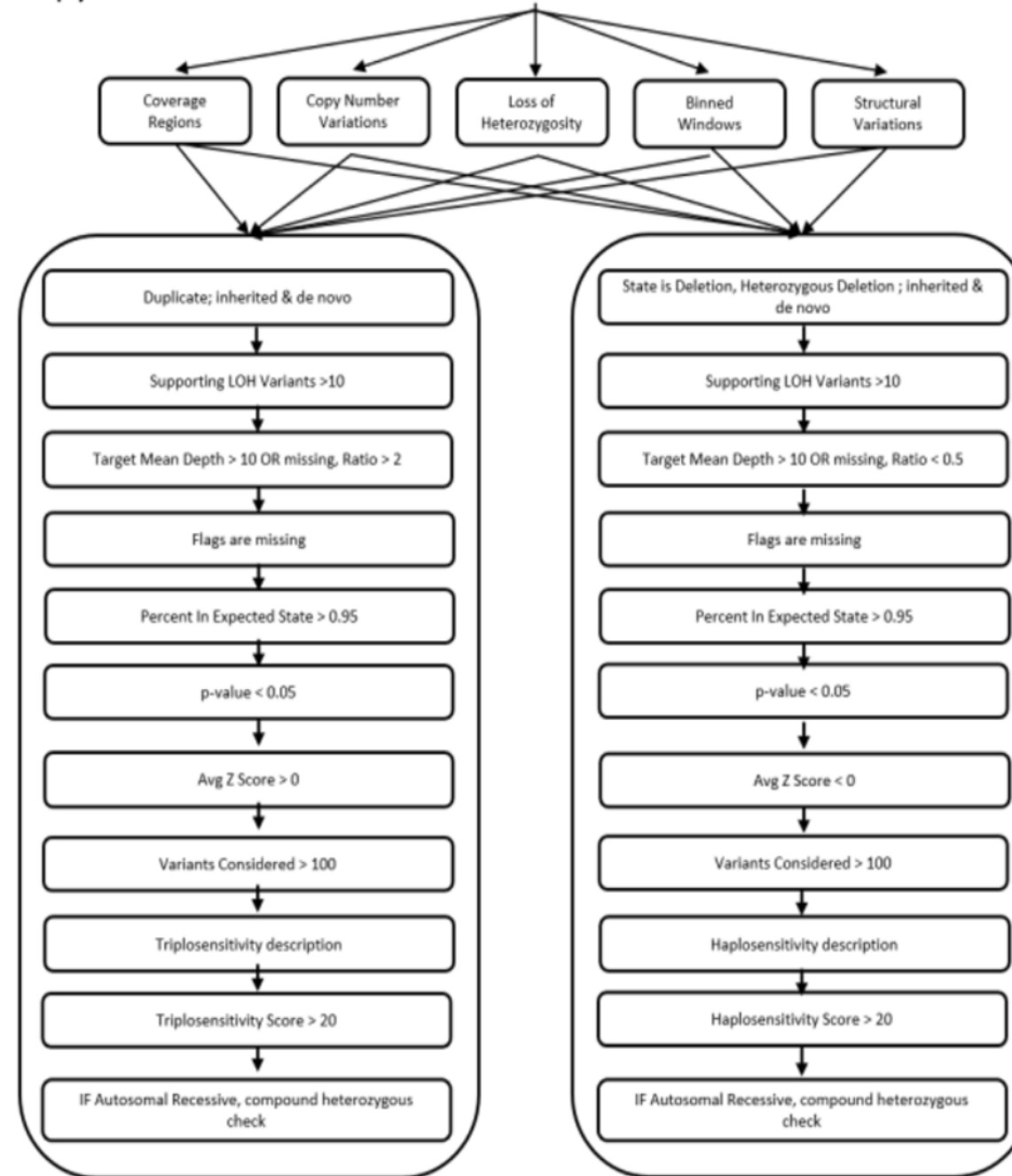
# Pipeline Part 3: Curation & Reporting

After QC checks and sample relatedness verification, the analysis branches into multiple preset categories for curation.



# Variant filtration

Copy number and structural variation filter: Raw VCF & BAM



# Variants pathogenicity annotation



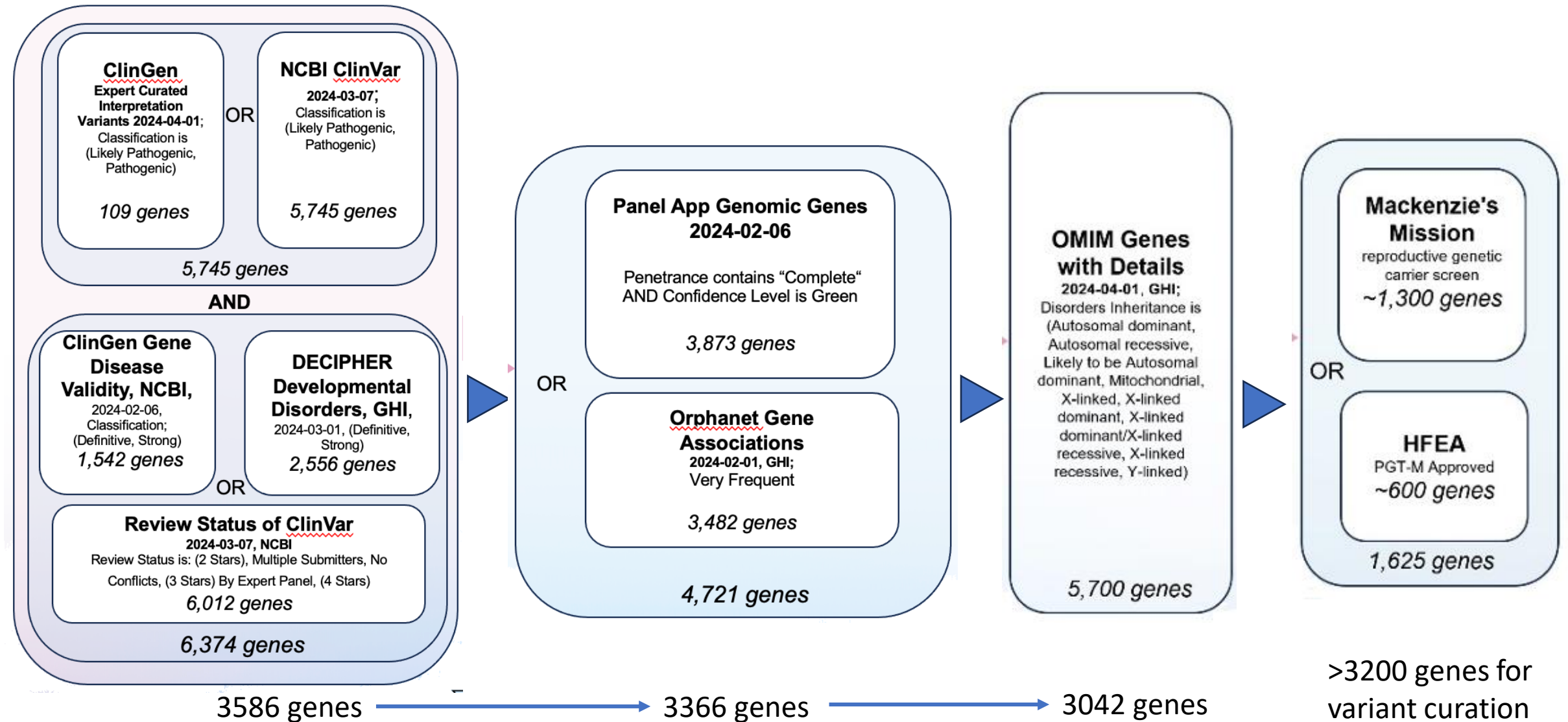
**Variants annotation** performed from >50 annotation databases, pathogenicity prediction algorithms, and ACMG guidelines.

*1kG Phase3 - Variant Frequencies 5a with Genotype Counts, GHI 1kG Phase3 CNVs and Large Variants 5b V2, GHI , CADD Scores InDels v1.5, UW, ClinGen Gene Disease Validity 2021-01-14, NCBI, ClinGen Gene Dosage Sensitivity 2022-02-03, NCBI , ClinGen Region Dosage Sensitivity 2022-01-07, NCBI , Clinical Genomic Database 2022-02-23, GHI, ClinVar 2022-01-06, NCBI , ClinVar Assessments 2022-01-06, NCBI ClinVar CNVs and Large Variants 2022-01-06, NCBI , ClinVar Transcript Counts 2022-01-06, NCBI, ClinVarCNVsandLargeVariant Assessments 2022-01-06, NCBI , Conservation Scores Exonic, GHI , dbSNP 155, NCBI , DECIPHER Developmental Disorders 2021-12-02, GHI, DECIPHER Population CNV v9.2DGV CNVs - Gold Standard Variants 2016-05-15 v3, DGV , Gene Identifiers and Descriptions 2021-11-30, GHI , Genetics Home Reference 2022-05-12, GHI, Genomic Super Dups 2014-10-19, UCSC gnomAD - Gene Constraint 2.1.1 v2, BROAD, gnomAD Exomes Variant Frequencies 2.0.1, BROAD , GnomAD High Frequency CNV Regions 2019-11-25, GHI , gnomAD Structural Variants 2.1, BROAD, Haploinsufficiency Predictions Version 3, DECIPHER, Human Phenotype Ontology 2022-02-21 , InterPro Regions 2019-09-18, GHI , Low Complexity Regions and Universal Mask-GHI, Missense Badness and MPC, BROAD, MONDO 2021-05-13, GHI, Mondo Gene Disease Association 2020-07-25, MI , Multiple Sequence Alignments of 100 Vertebrates, UCSC, OMIM Genes 2022-02-01, GHI, Orphanet Gene Associations 2022-02-01, GHI, Reference Sequence GRCH37 g1k, 1000Genomes, Reference Sequence GRCh38, NCBI, RefSeq Genes 109.20211119, NCBI, Repeating Elements by RepeatMasker, UCSC , SIFT and PolyPhen2 Missense Predictions 2021-04-21, GHI*

# Gene selection criteria

- **Pathogenic** or Likely Pathogenic
- >90% certainty of being **disease-causing**
- **several submissions** with no conflicts between submissions
- High to **complete penetrance** (>80% of individuals develop the disease)
- Simple (**monogenic**) mode of inheritance:
  - Initially only Autosomal dominant or recessive, X-linked, Mitochondrial
- **No cure** available
- **Severe** phenotype

# 3200 genes selected



# Gene Selection

- **3,200 genes** included as the standard test
- **Can include any other on demand**

## Candidate Genes

This gene list includes 3246 genes

A

AAAS NCBI: 8086	AAGAB NCBI: 79719	AARS1 NCBI: 16	AARS2 NCBI: 57505	AASS NCBI: 10157	ABAT NCBI: 18	ABCA1 NCBI: 19
ABCA12 NCBI: 26154	ABCA2 NCBI: 20	ABCA3 NCBI: 21	ABCA4 NCBI: 24	ABCB11 NCBI: 8647	ABCB4 NCBI: 5244	ABCB6 NCBI: 10058
ABCB7 NCBI: 22	ABCC6 NCBI: 368	ABCC8 NCBI: 6833	ABCC9 NCBI: 10060	ABCD1 NCBI: 215	ABCD4 NCBI: 5826	ABCG5 NCBI: 64240
ABCG8 NCBI: 64241	ABHD12 NCBI: 26090	ABHD5 NCBI: 51099	ABL1 NCBI: 25	ACAD8 NCBI: 27034	ACAD9 NCBI: 28976	
ACADM NCBI: 34	ACADS NCBI: 35	ACADSB NCBI: 36	ACADVL NCBI: 37	ACAN NCBI: 176	ACAT1 NCBI: 38	ACBD5 NCBI: 91452
ACE NCBI: 1636	ACO2 NCBI: 50	ACOX1 NCBI: 51	ACP4 NCBI: 93650	ACP5 NCBI: 54	ACSF3 NCBI: 197322	ACSL4 NCBI: 2182
ACTA1 NCBI: 58	ACTA2 NCBI: 59	ACTB NCBI: 60	ACTC1 NCBI: 70	ACTG1 NCBI: 71	ACTG2 NCBI: 72	ACTL6B NCBI: 51412
ACTN1 NCBI: 87	ACTN2 NCBI: 88	ACTN4 NCBI: 81	ACVR1 NCBI: 90	ACVRL1 NCBI: 94	ACY1 NCBI: 95	ADA NCBI: 100
ADA2 NCBI: 51816	ADAM17 NCBI: 6868	ADAM22 NCBI: 53616	ADAM9 NCBI: 8754	ADAMTS10 NCBI: 81794	ADAMTS13 NCBI: 11093	
ADAMTS17 NCBI: 170691	ADAMTS18 NCBI: 170692	ADAMTS2 NCBI: 9509	ADAMTSL2 NCBI: 9719	ADAMTSL4 NCBI: 54507	ADAR NCBI: 103	
ADAT3	ADCV5	ADCV6	ADCBG1	ADCBG6	ADCBV1	ADK

Back

Next

# **validation method using GIAB**

# validation method



- Flow sorted cells from several Genome-in-a-bottle (GIAB vs 3.3.2): NA12878, NA12877, ..
- Compared genomic DNA to amplified DNA (using RepliG)
- CNV & ploidy were assessed via transmission & read binning for Copy Number
- Tandem repeat disorders were reported via carrier repeat number
- De Novo mutation accuracy was calculated after the filtering but BEFORE annotation.
- Annotation significantly improved the accuracy.
- *De novo* mutations after annotation were reconfirmed by Sanger.

# validation results using GIAB

## Mutation performance metrics prior to annotation (SNVs)

<b>Specificity</b>	<b>99.998%</b>
<b>Accuracy</b>	<b>99.997%</b>
<b>Sensitivity</b>	<b>92.2%</b>
<b>Precision</b>	<b>98.0%</b>
<b>Negative Predictive Value</b>	<b>99.992%</b>
<b>False Positive Rate</b>	<b>0.0023%</b>
<b>False Negative Rate</b>	<b>2.93%</b>

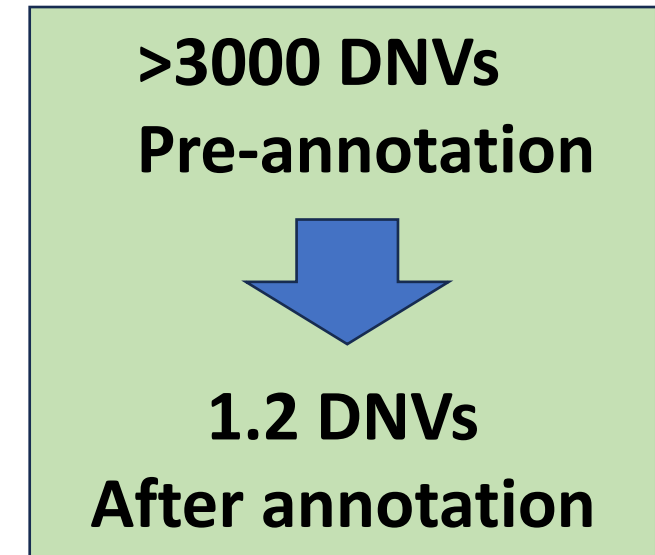
# Rate of Pathogenic *de novo* mutations

**Pre-annotation** detection of the novo mutations:  
3,632 DNVs in NA12878 5-cells model **biopsies**

**Post-annotation** detection of the novo mutations:  
Average of **1.17 pathogenic DNVs** per sample

**Further confirmation pre-reporting:**

- **Sanger** re-testing for DNV prior to reporting
- Re-confirmation using **Spent culture media**



**validation method using  
rebiopsy vs whole embryo**

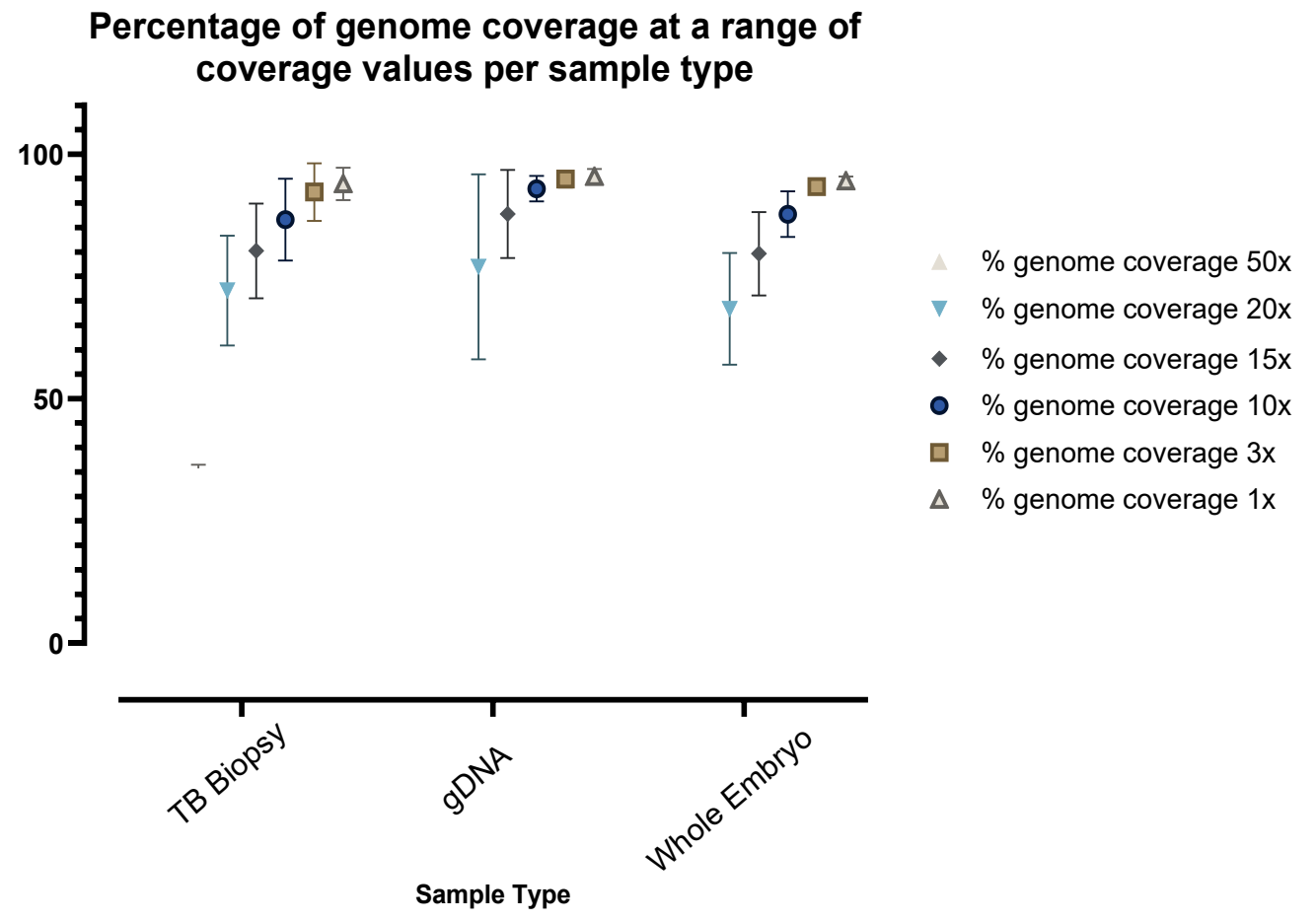
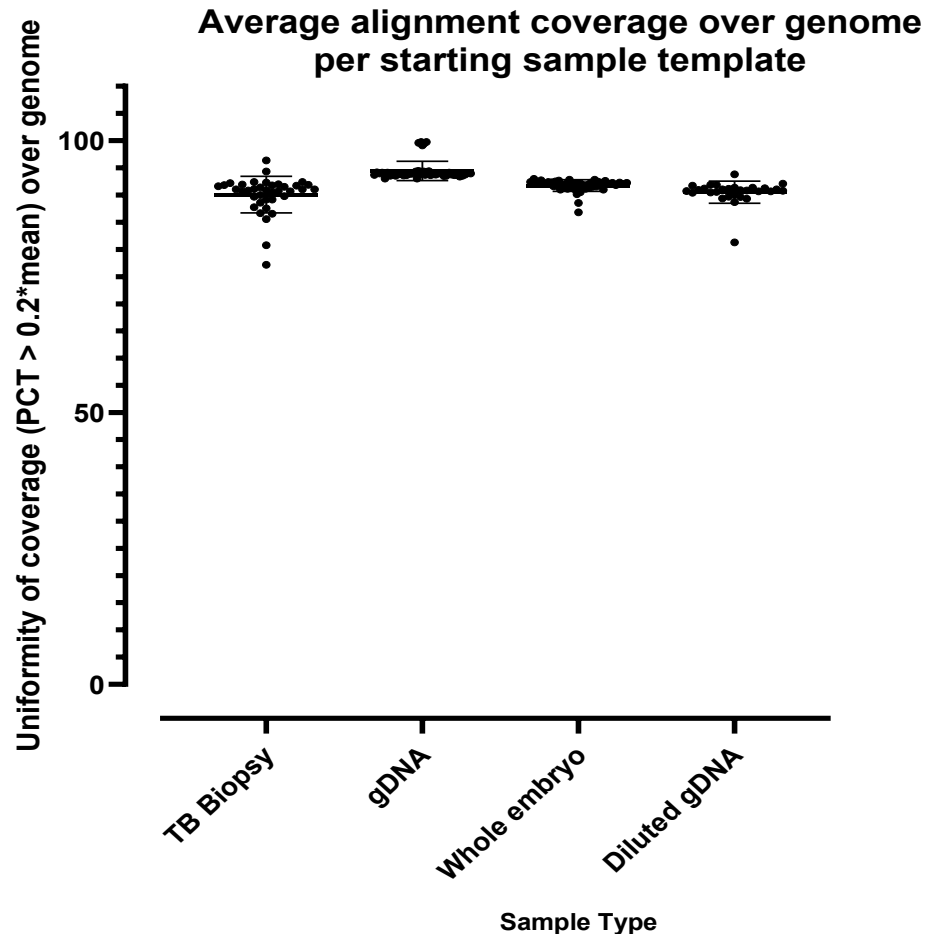
# Clinical validation study (NCT05739890):



- **Patients:** - 100 carrier couples of monogenic diseases undergoing PGT-M
- **Procedure:** - Embryo biopsy and MDA amplification
  - PGT-WGS at x40 for embryos and parents at x30
  - Affected embryos re-biopsied
- **Aim:** - Compare PGT-WGS of TE biopsy (query) vs. whole embryo (Truth).
  - Determine at what paternal age is WGS more indicated
- **Metrix:** - genome in a bottle benchmarking standards (**recall, precision, F1**)

# Clinical Validation: uniform coverage

Uniform coverage was very similar between TB, whole embryo and gDNA



# Clinical validation results:

Whole embryo	biopsy WGS	Biopsy PGT-M	whole Embryo WGS	biopsy PGT-WGS aneuploidy	Biopsy PGT-A	#
affected	affected	affected	euploid	euploid	euploid	16
affected	affected	affected	euploid	euploid	NR	4
affected	affected	affected	euploid	euploid	chaotic	1
affected	affected	affected	Aneuploid	Aneuploid	Aneuploid	8
affected	affected	affected	47XX,+3	47XX,+3	47XX,+3, mos -1	1
affected	affected	affected	45XX,-19	45XX,-19, -5q	46XX,-19, +21	1
affected	affected	affected	Complex	complex	complex	3
affected	affected	affected	69,XXY	69,XXY	complex	1
affected	affected	affected	45XX,-18	44XX,-18, -9	NR	1
<b>TOTAL</b>	<b>100%</b>			<b>95%</b>		<b>36</b>

Kahraman, Cetinkaya, Munné, Murphy (submitted)

# Clinical Validation Results:

## Pathogenic *de novo* Variants

**195 candidate mutations**  
detected in 22 embryos

**Filter applied:** Allele freq thresholding, depth >3 reads, Map Quality >20, pop. freq <10%, high severity, de novo, disease associated genes; SNVs & indels known variants only.

**4 De novo mutations**  
selected for confirmation

DMD	rs104894797	Duchenne muscular dystrophy
TRMU	rs779022860	Liver Failure, Infantile
LOX	rs1754538399	Aortic aneurysm, familial thoracic
EYA4	rs1554275988	Deafness, autosomal dominant

**2 De novo mutations confirmed in the whole embryo**

**9% embryos have confirmed *de novo* mutations (2/22)**

# ***Genetic counseling***

# Advantage over other PGT-M platforms

- Embryos classified as heterozygous for a recessive disease **may still carry a de novo** mutation and be affected.
- PGT-WGS can detect other **inherited** or **de novo** mutation in other genes not tested for the PGT-M indication.
- **Initial indications:**
  - PGT-M
  - Advanced paternal age
  - Consanguinity

# Genetic counseling and reporting



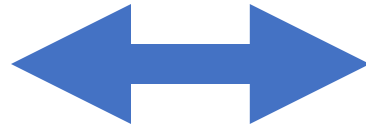
Two approaches:

- **Report only pathogenic:**
  - **Only pathogenic** or likely pathogenic variants are reported following the prenatal diagnosis approach to WGS
  - This is **more conductive** to current PGT reporting in IVF centers which have limited time and capabilities for genetic counseling.
- **Report other variants and traits** (i.e. PGT-P for IQ):
  - Some demanding parents may want **extensive counseling** on each embryo, each gene and each trait requiring specialized services beyond regular IVF set ups

# Conclusions



- It took **30 years** (1993 – 2023) to go from the first PGT-A (XY,13,18,21) to PGT-WGS of embryos
- **30% of euploid embryos still do not implant. Metabolomics and WGS** may identify others, while 10% loss may be due to the embryo transfer process itself.
- Although there is a variety of platforms that can perform **comprehensive PGT for –A –M –SR –P**, only **WGS** can detect **de novo mutations PLUS** everything else.



GenEmbryomics PGT-WGS test is distributed by Progenesis

# Credits to:

## GenEmbryomics:

- Nick Murphy, PhD
- Kyle Day, PhD
- Shannon Wieloch, PhD
- Kim Skellington, PhD
- Vanessa Cortes, PhD

## Memorial Sisli Hospital:

- Prof. Semra Kahraman, M.D
- Murat Cetinkaya, M.D., PhD

## Questions?

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